



Roberto Neira Roa

Profesor Emérito de la Universidad de Chile

6 de Noviembre del 2025

Edición Genética: nueva herramienta para enfrentar los desafíos en Sanidad Animal

José M. Yáñez

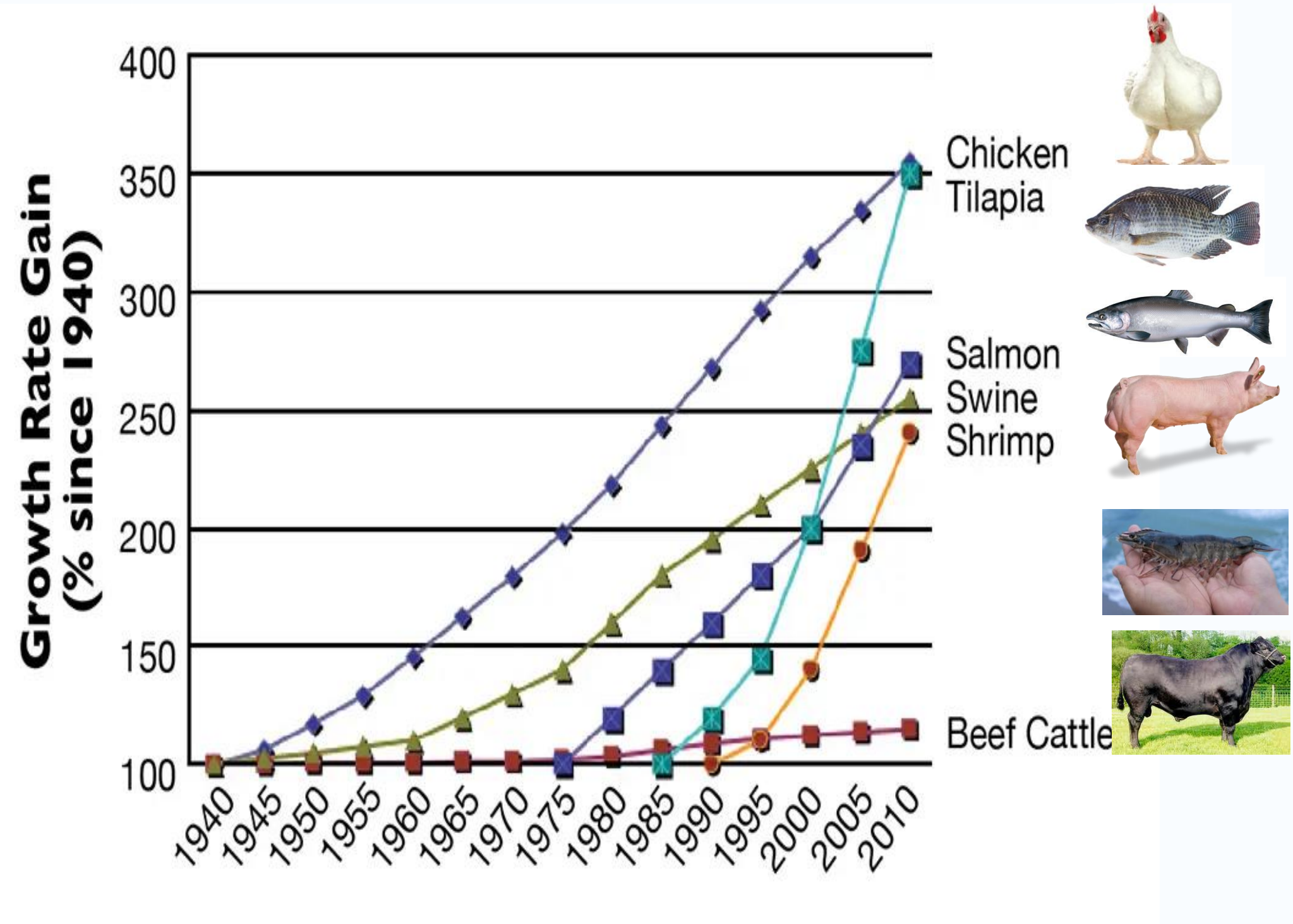
Facultad de Ciencias Veterinarias y Pecuarias
Universidad de Chile



MEJORAMIENTO GENÉTICO

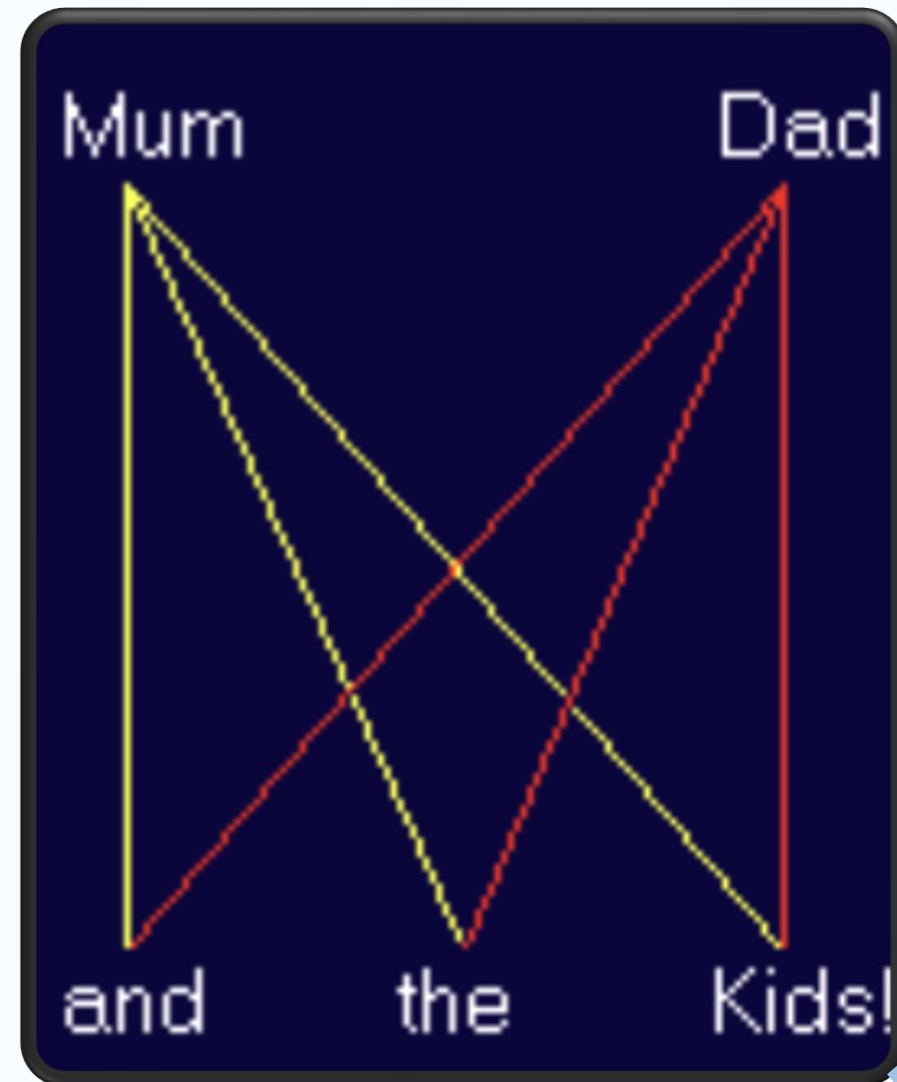
Pilar fundamental para incrementar eficiencia y sostenibilidad de la industria

- ✓ Acelera tasa de crecimiento
- ✓ Incrementa resistencia a enfermedades y sobrevivencia
- ✓ Mayor calidad y rendimiento de producto

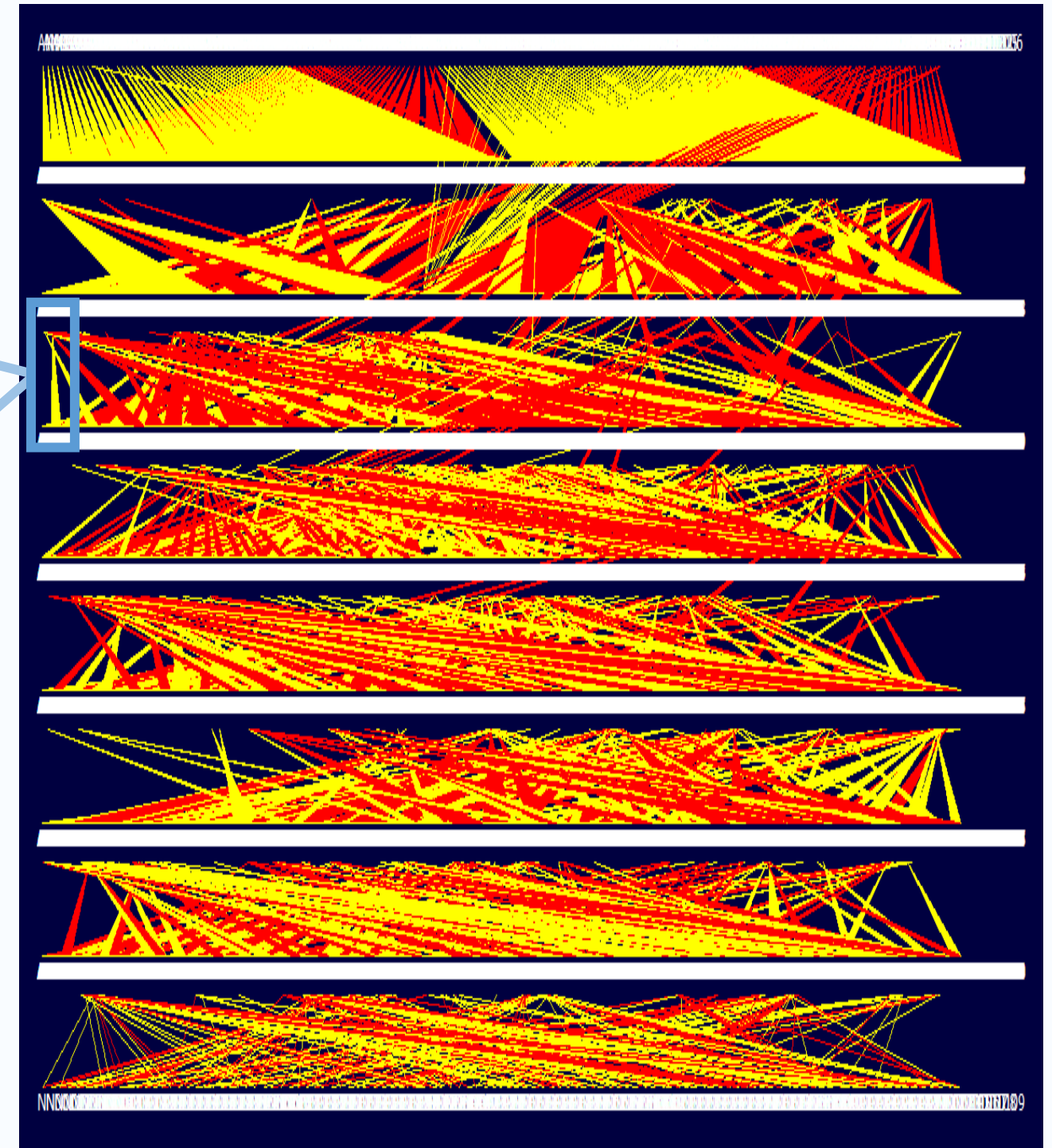


PREDICCIÓN DEL MÉRITO GENÉTICO

1. PEDIGRÍ

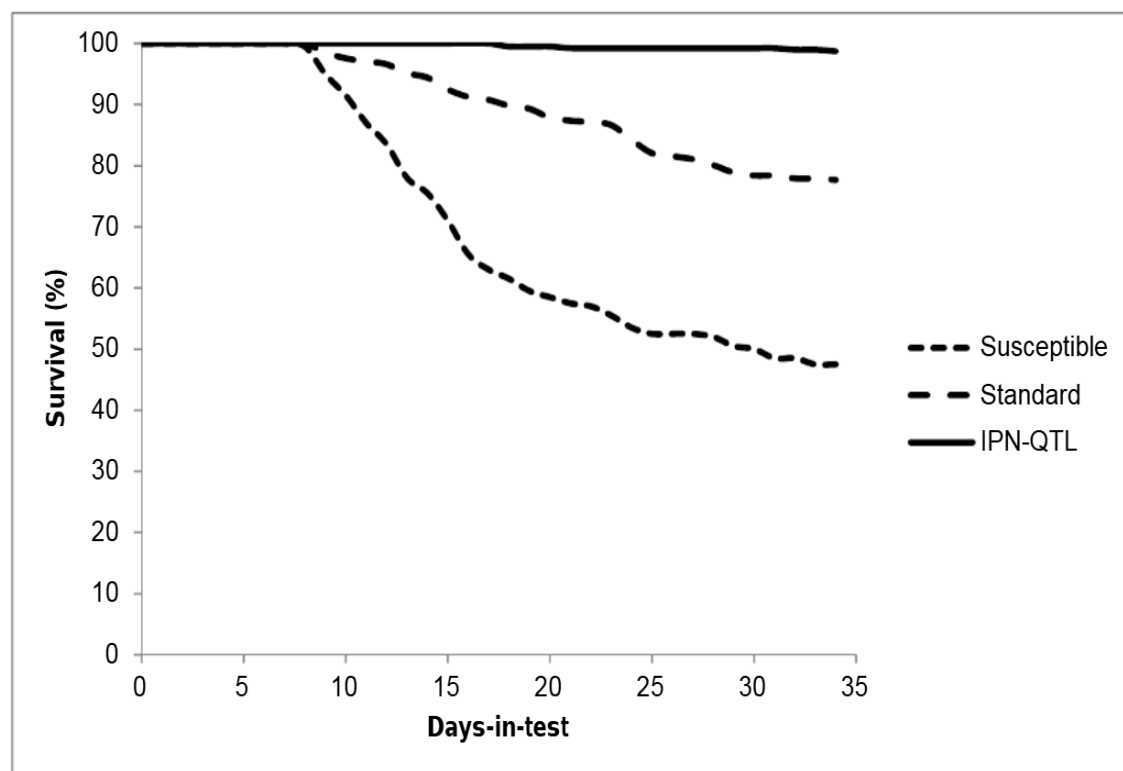
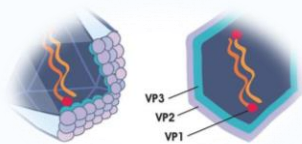


Genealogía salmón coho



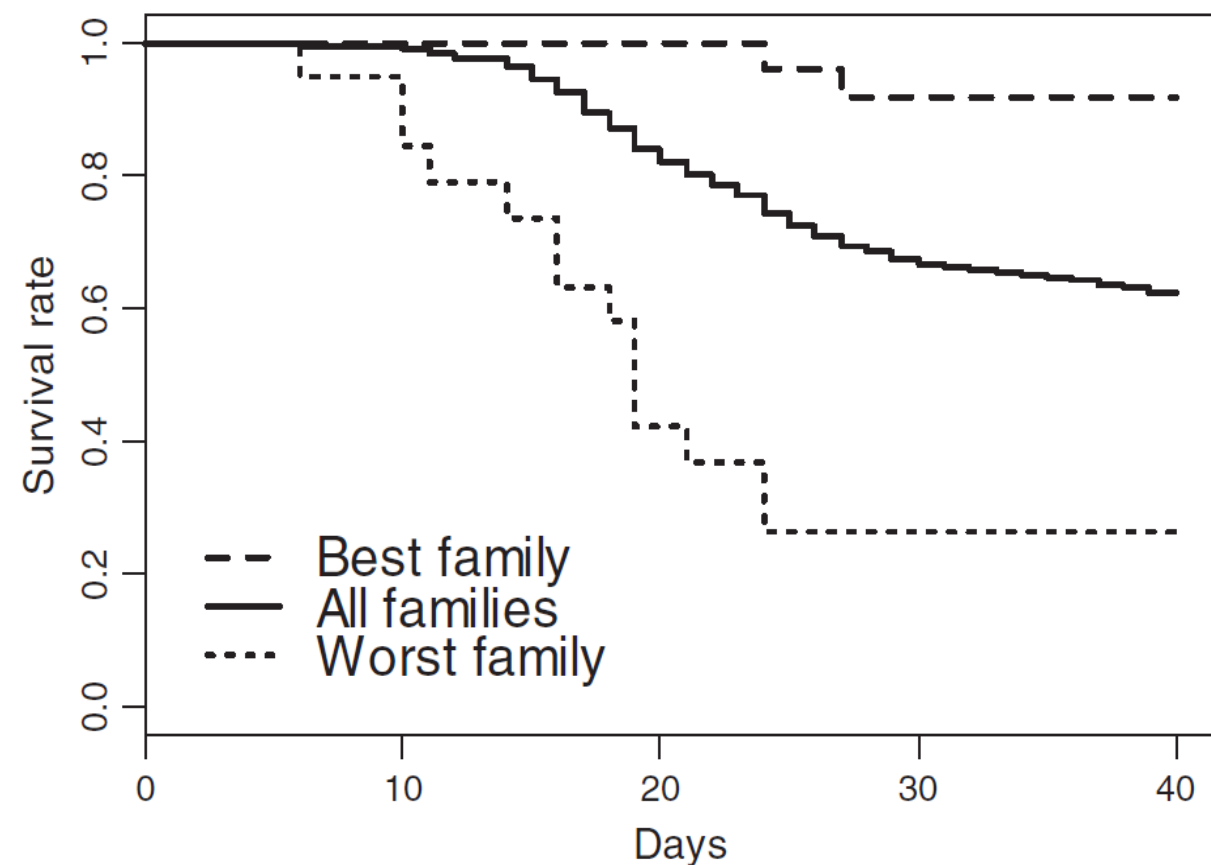
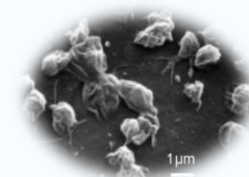
2. FENOTIPOS

IPN



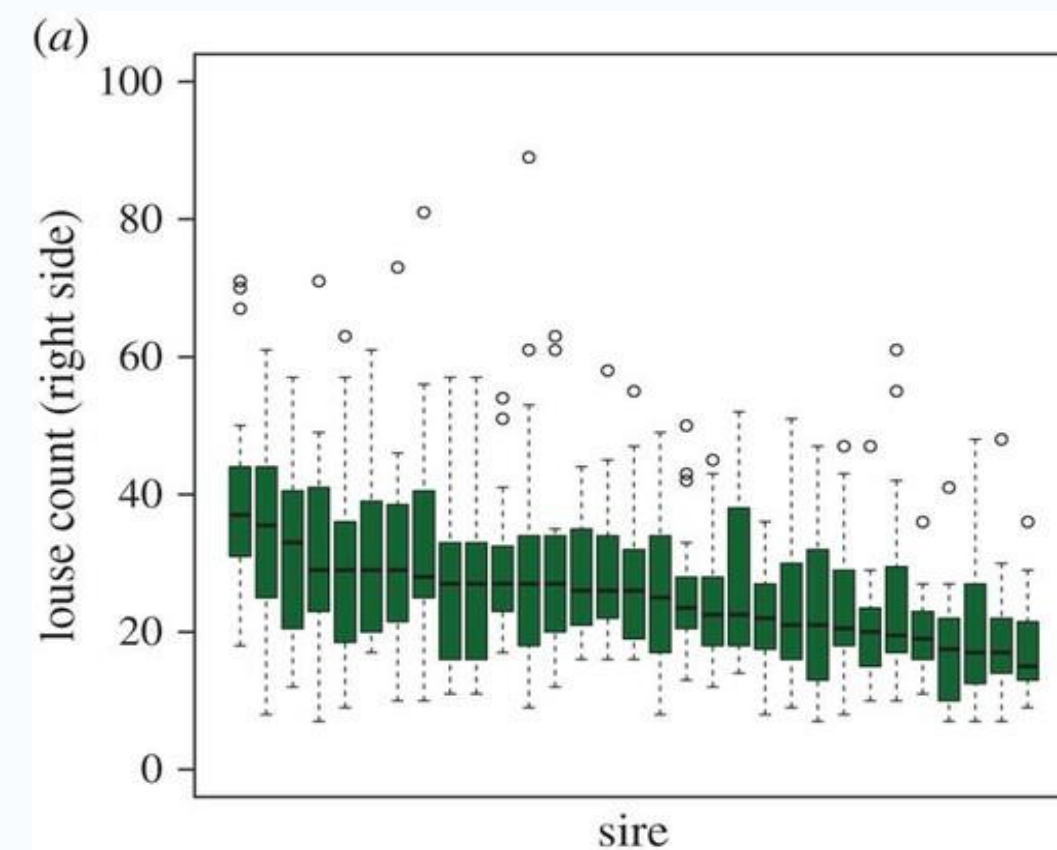
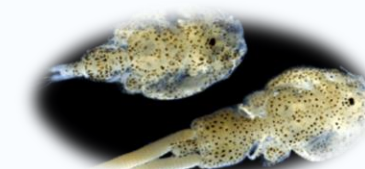
Moen et al. 2015

SRS



Yáñez et al. 2013

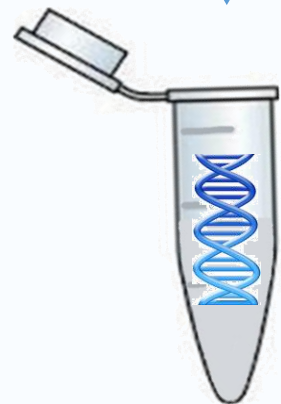
Sea lice



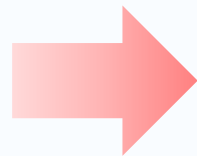
Gharbi et al. 2015

3. GENOTIPOS

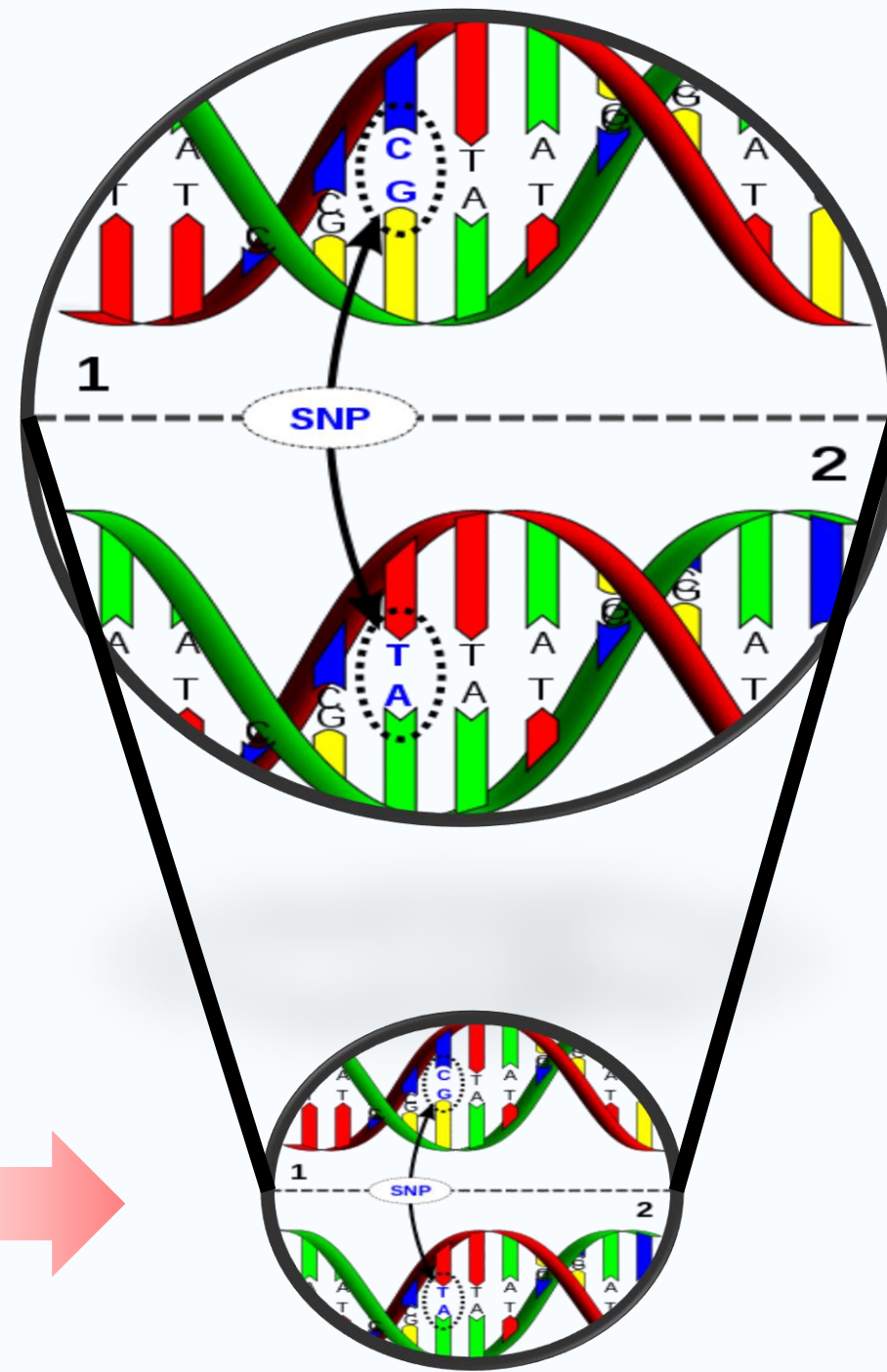
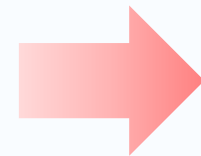
SINGLE NUCLEOTIDE POLYMORPHISMS (SNP)



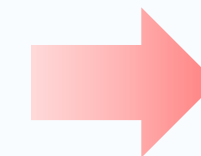
Purificación de DNA



Secuenciación

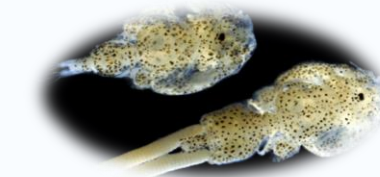


Descubrimiento de SNPs



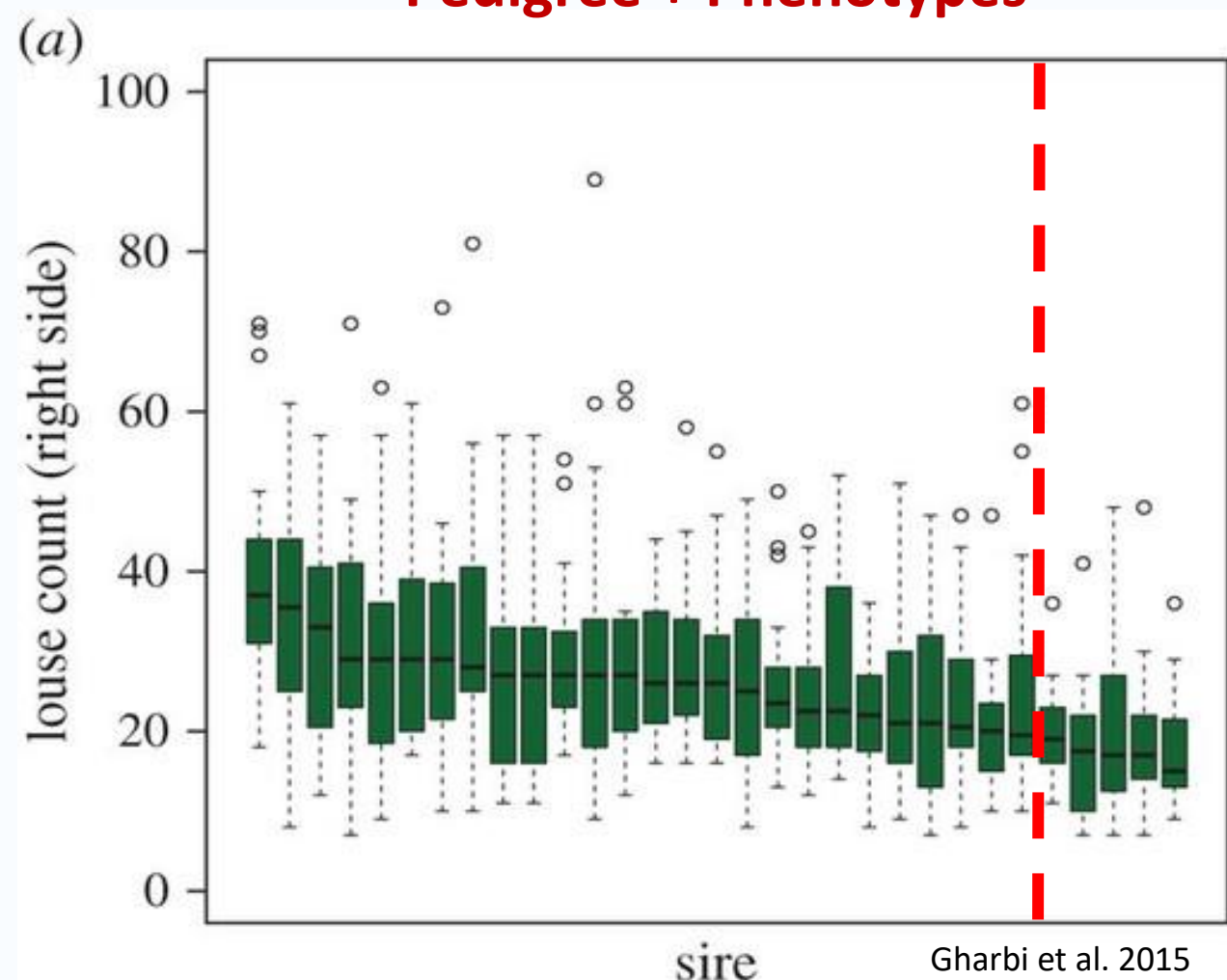
Genotipado de alto rendimiento

TRADITIONAL SELECTION VS GENOMICS

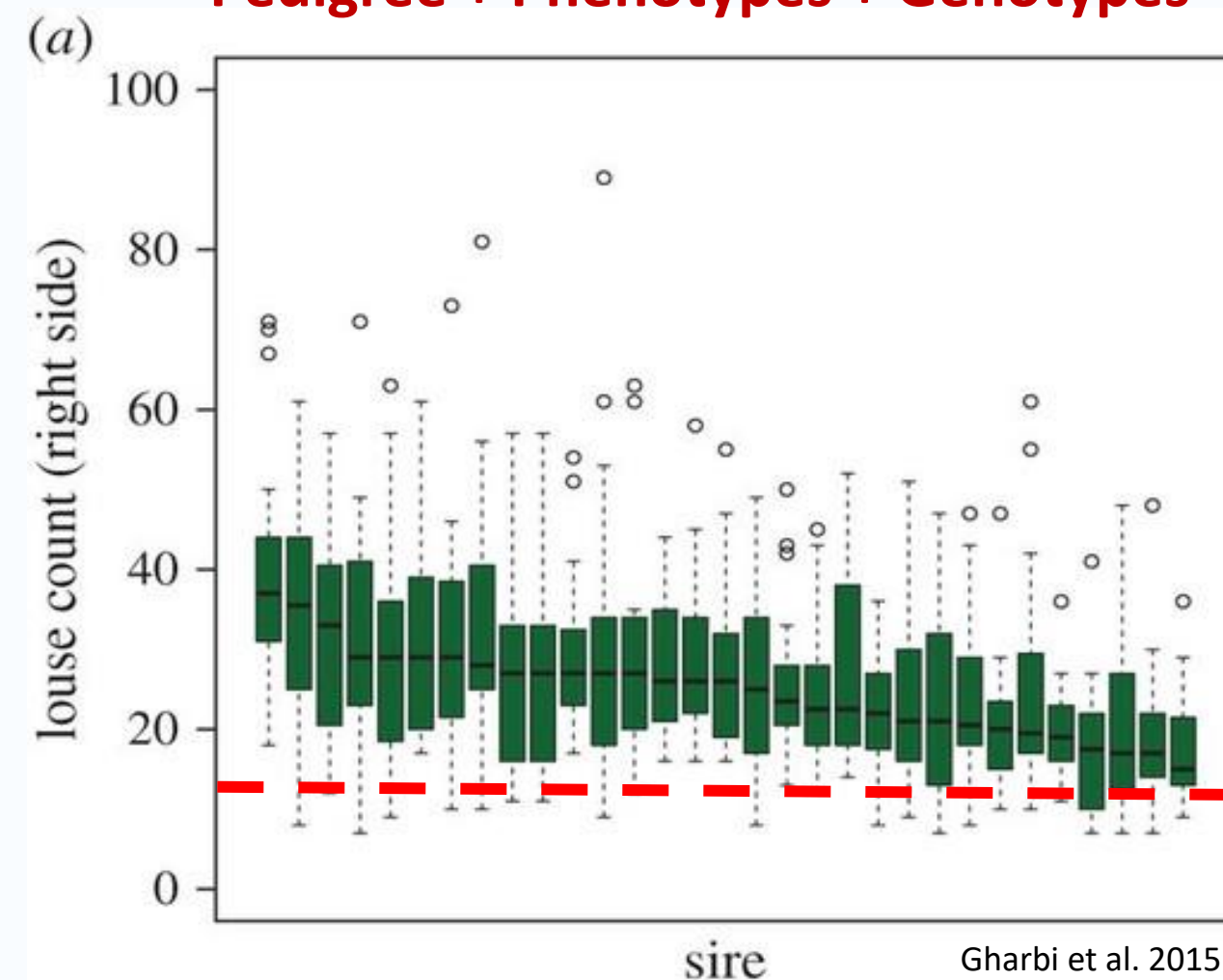


Sea lice

Pedigree + Phenotypes



Pedigree + Phenotypes + Genotypes



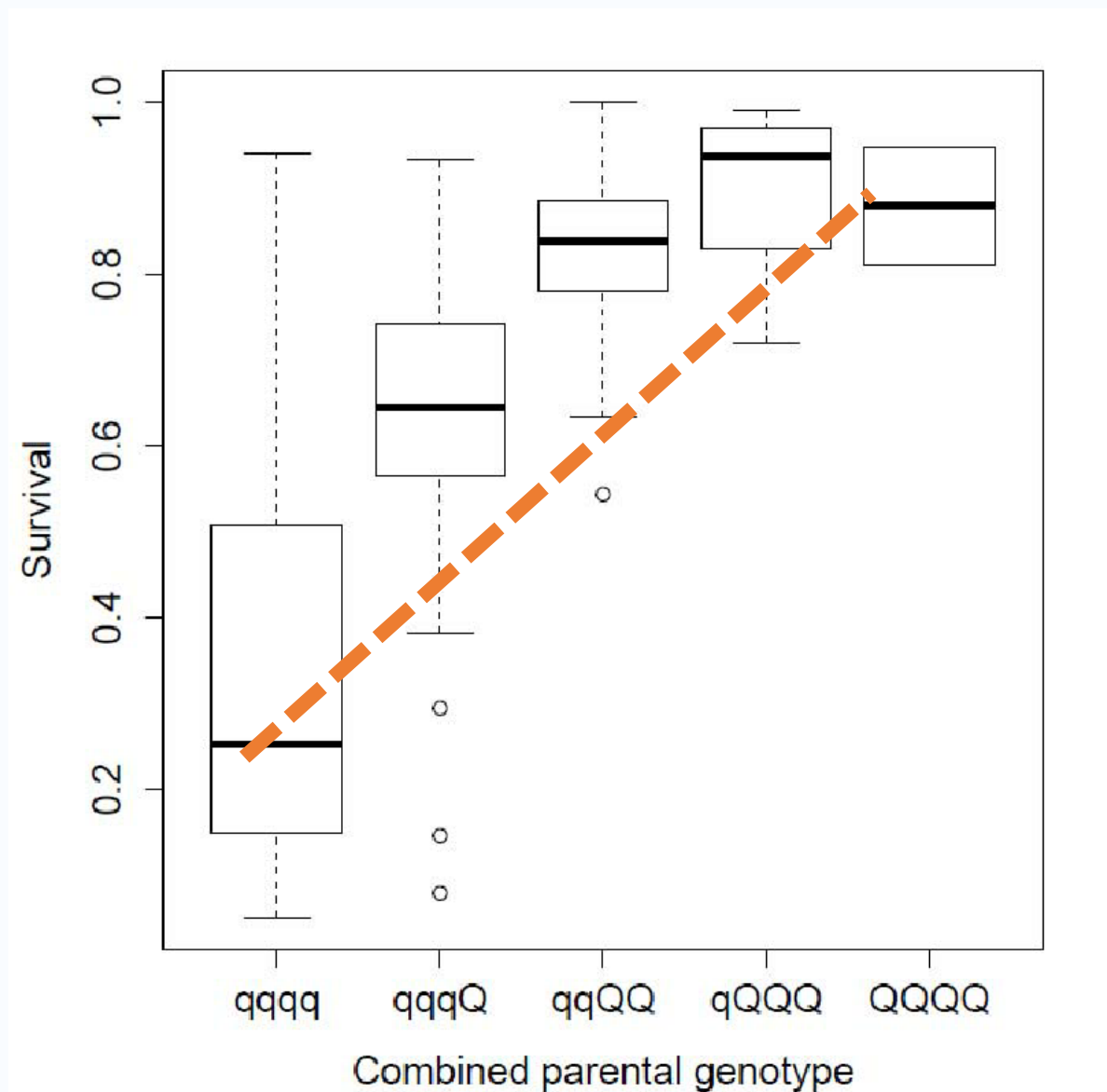
Selection of most resistant **families**

- **Low accuracy:** exploits only within-family variation
- **Low selection intensity:** predisposes to higher inbreeding

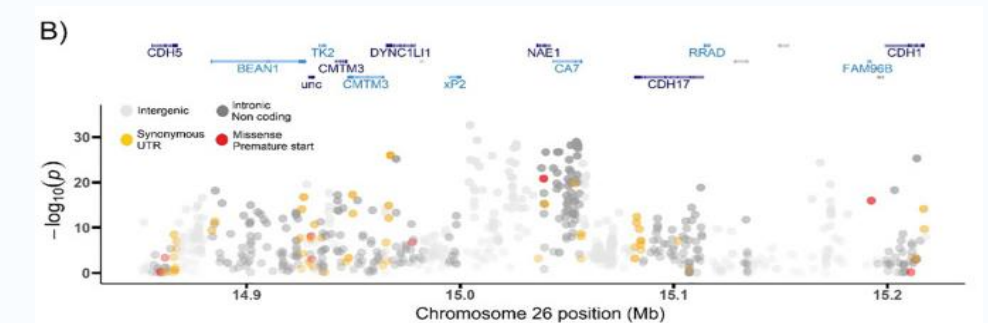
Selection of most resistant **individuals**

- **High accuracy:** exploits within- and between family variation
- **High selection intensity:** lower inbreeding levels

GENOME-WIDE ASSOCIATION STUDIES (GWAS) & QUANTITATIVE TRAIT LOCI (QTL)



***p-value* = 0.000000000000000000000000000000005**

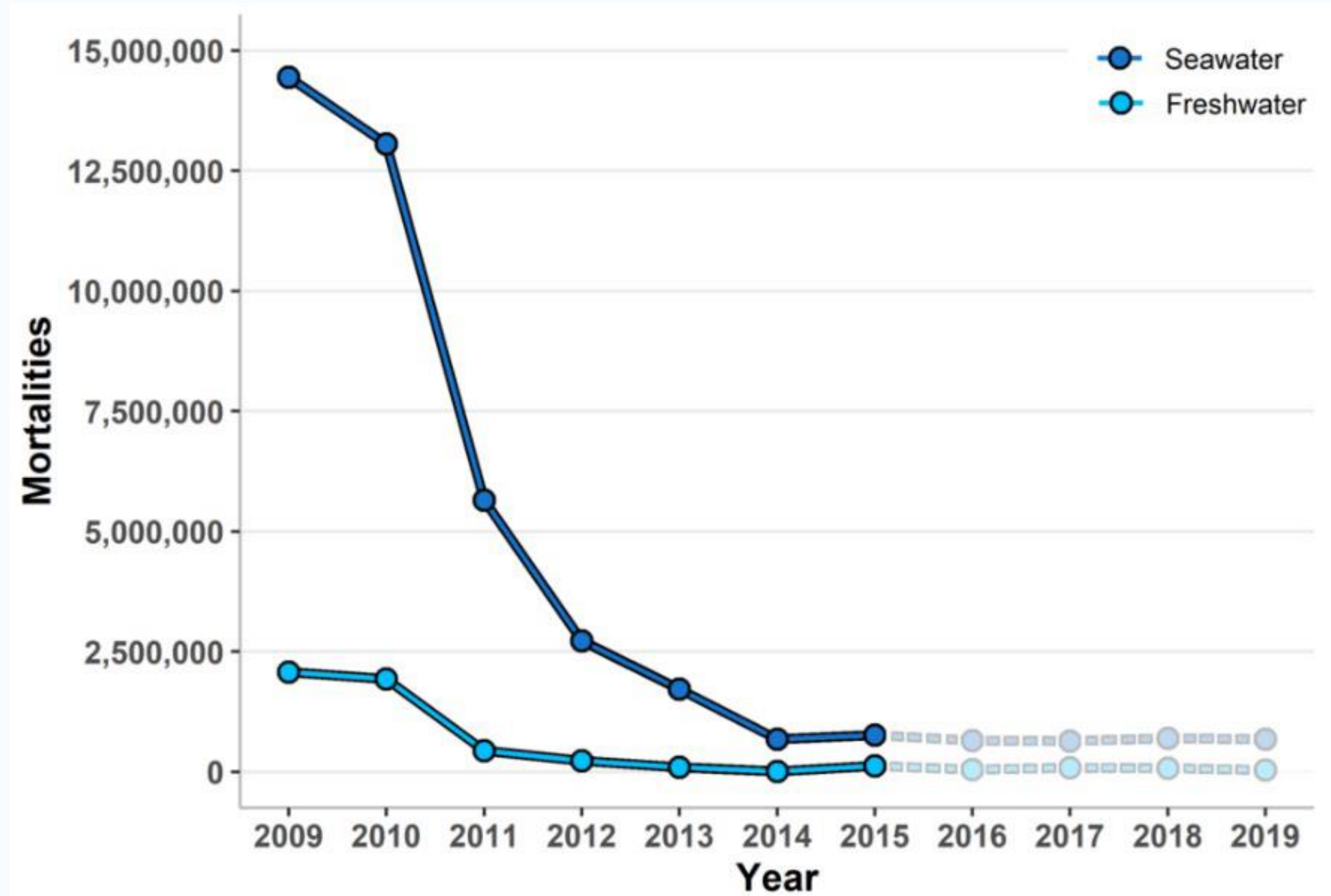


Manhattan Plot

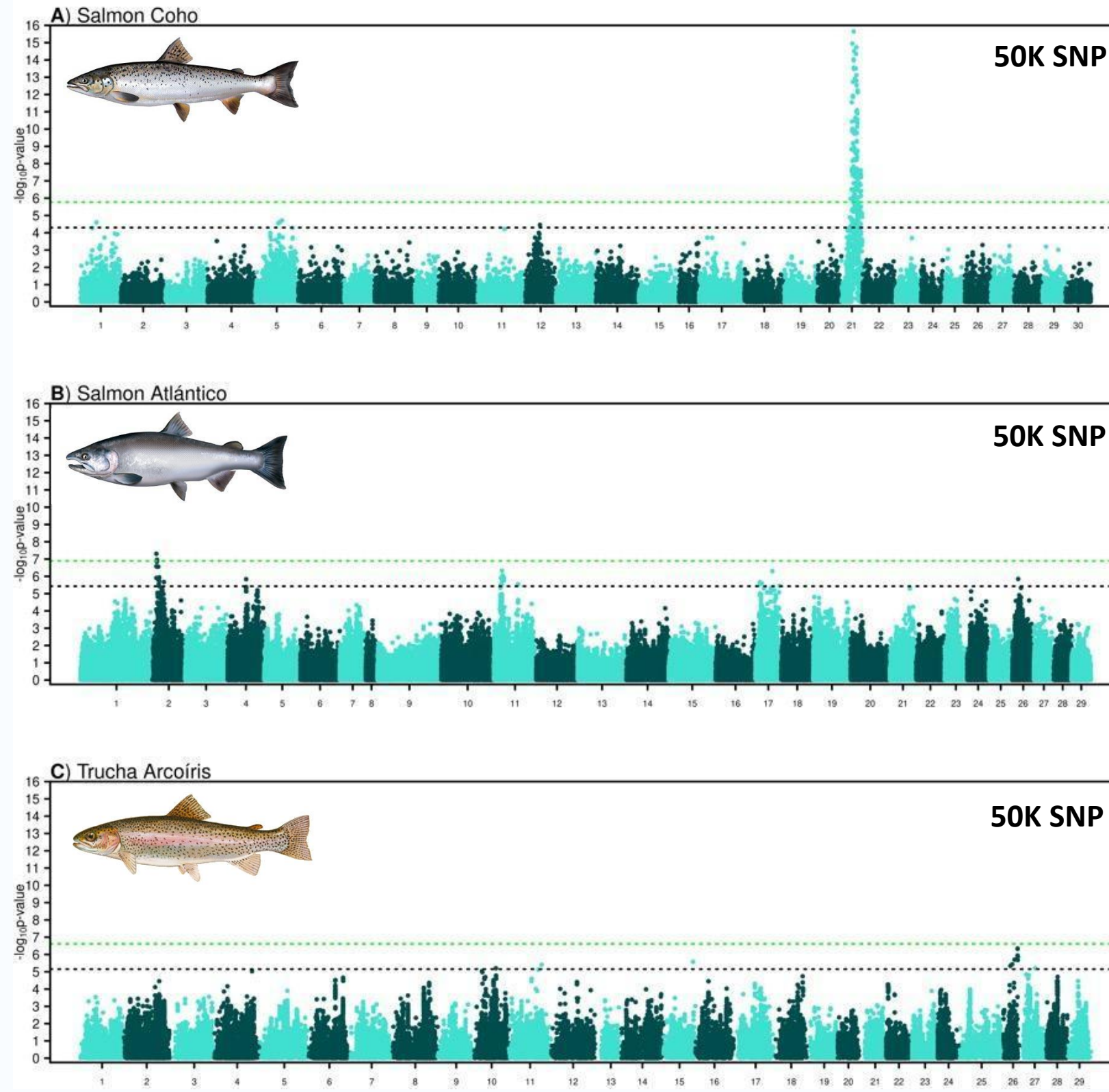
IPN Resistance in Atlantic salmon



QTL FOR IPN VIRUS RESISTANCE



GWAS SRS RESISTANCE (50K SNP)

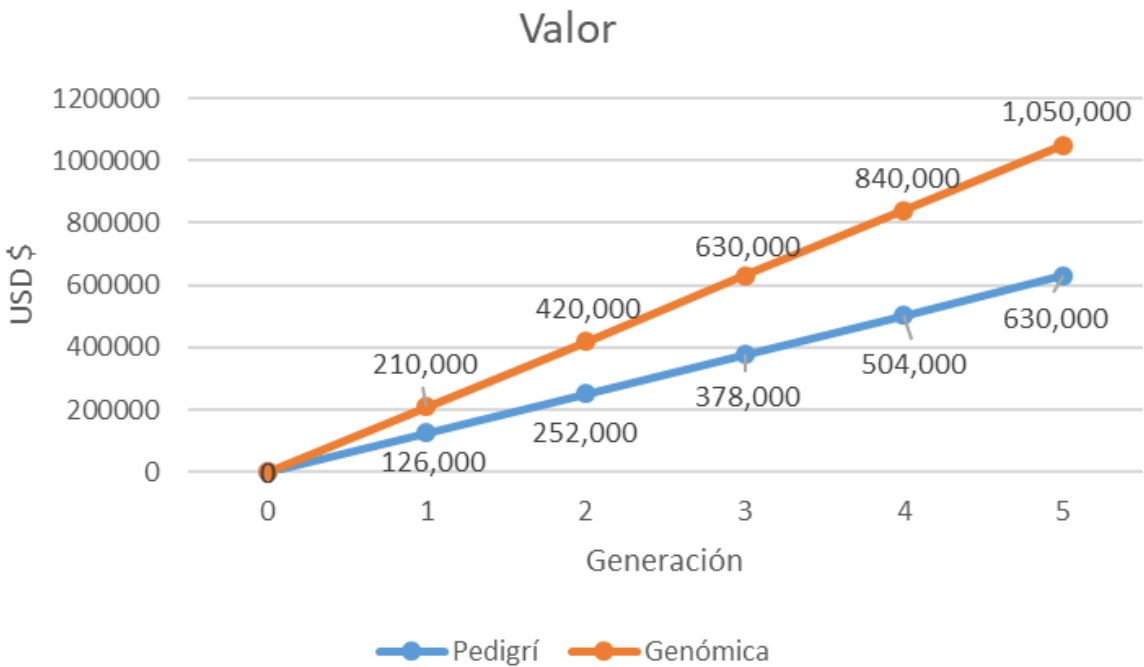
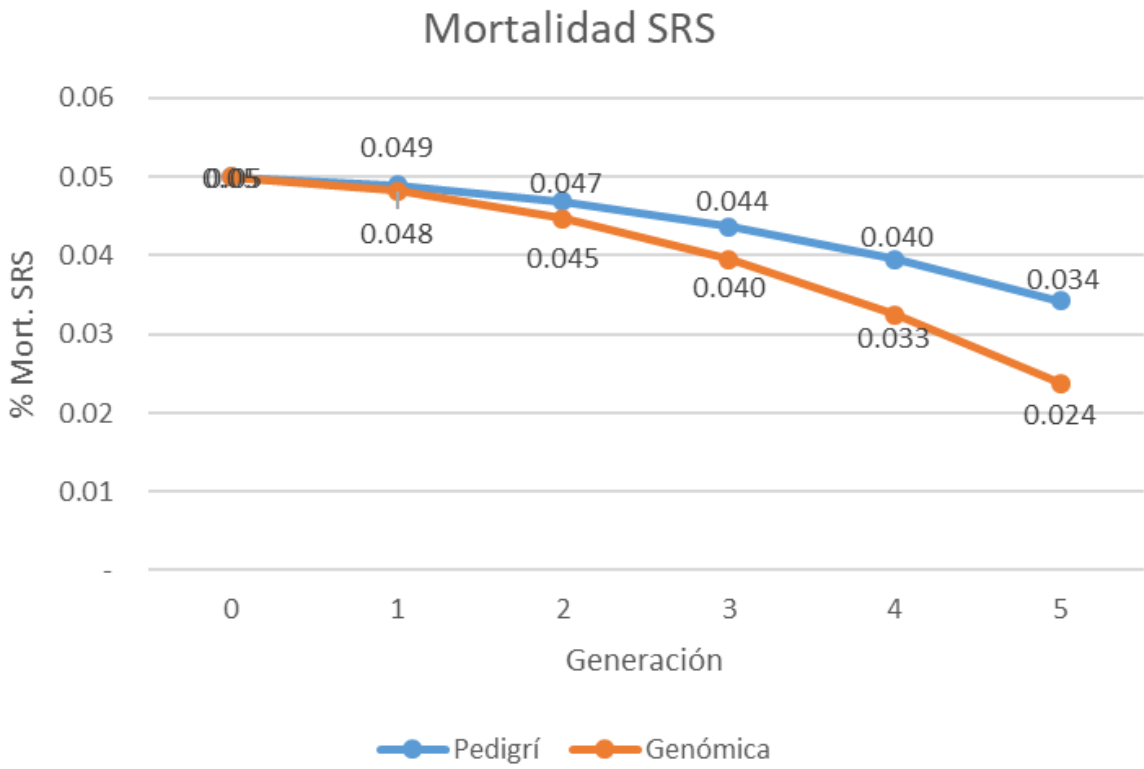
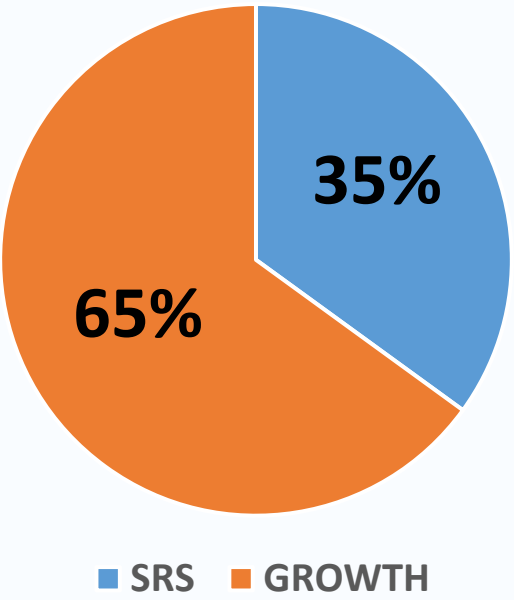


EXPECTED SELECTION RESPONSE

Assumptions

Item	Valor
Biomasa anual (tons)	10,000
Precio (\$ USD/k)	6
Mort. total (%)	10
Mort. SRS (%)	5
Perdidas Mort. (tons)	1,000
Perdidas Mort. (\$ USD)	12,000,000
Perdidas SRS (tons)	500
Perdidas SRS (\$ USD)	6,000,000

Selection Index

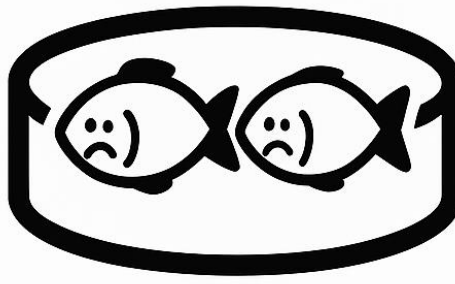
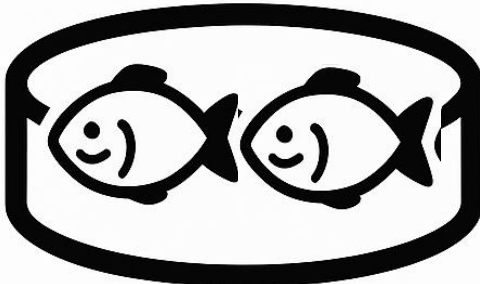
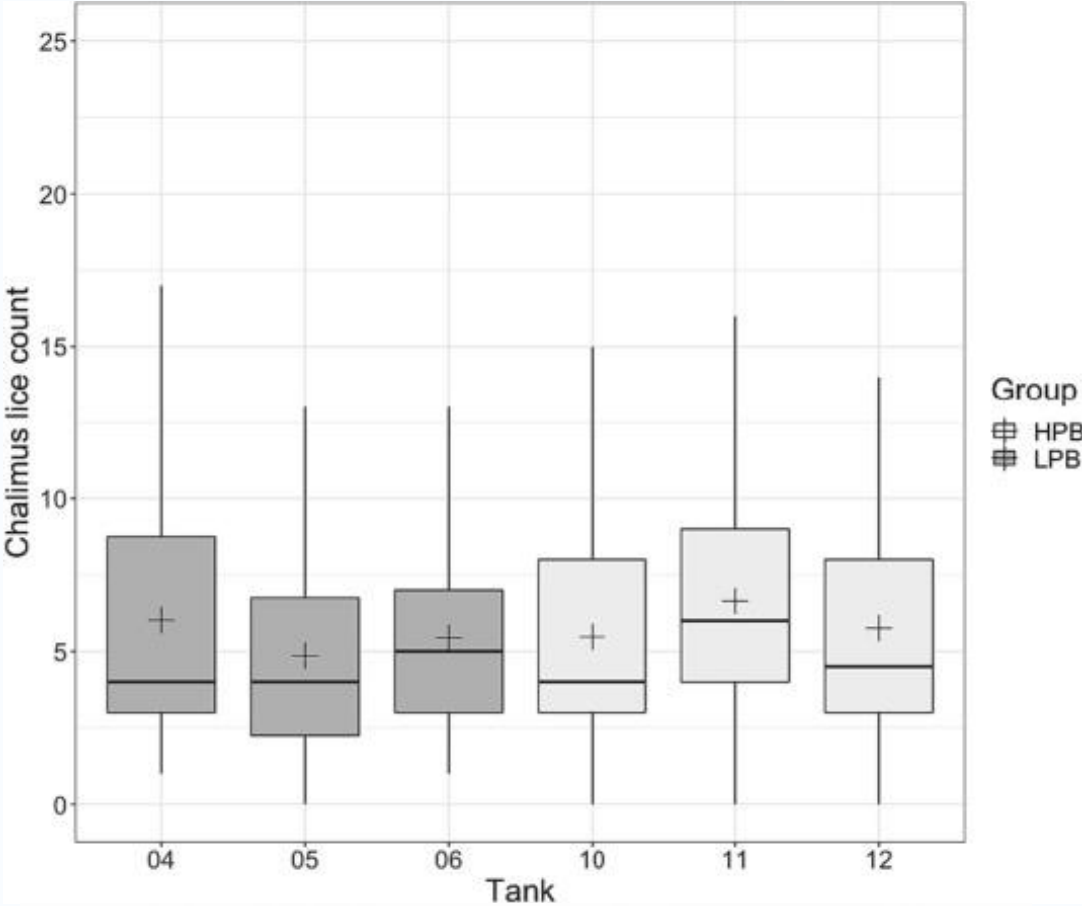
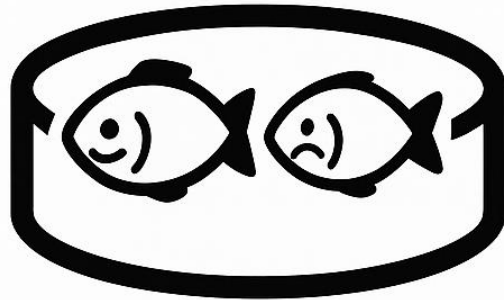
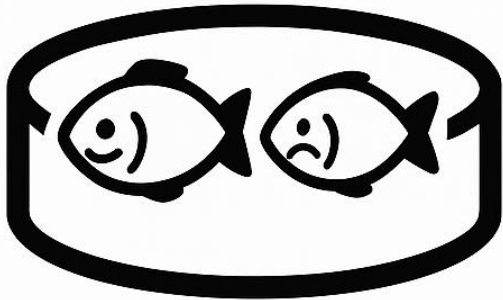
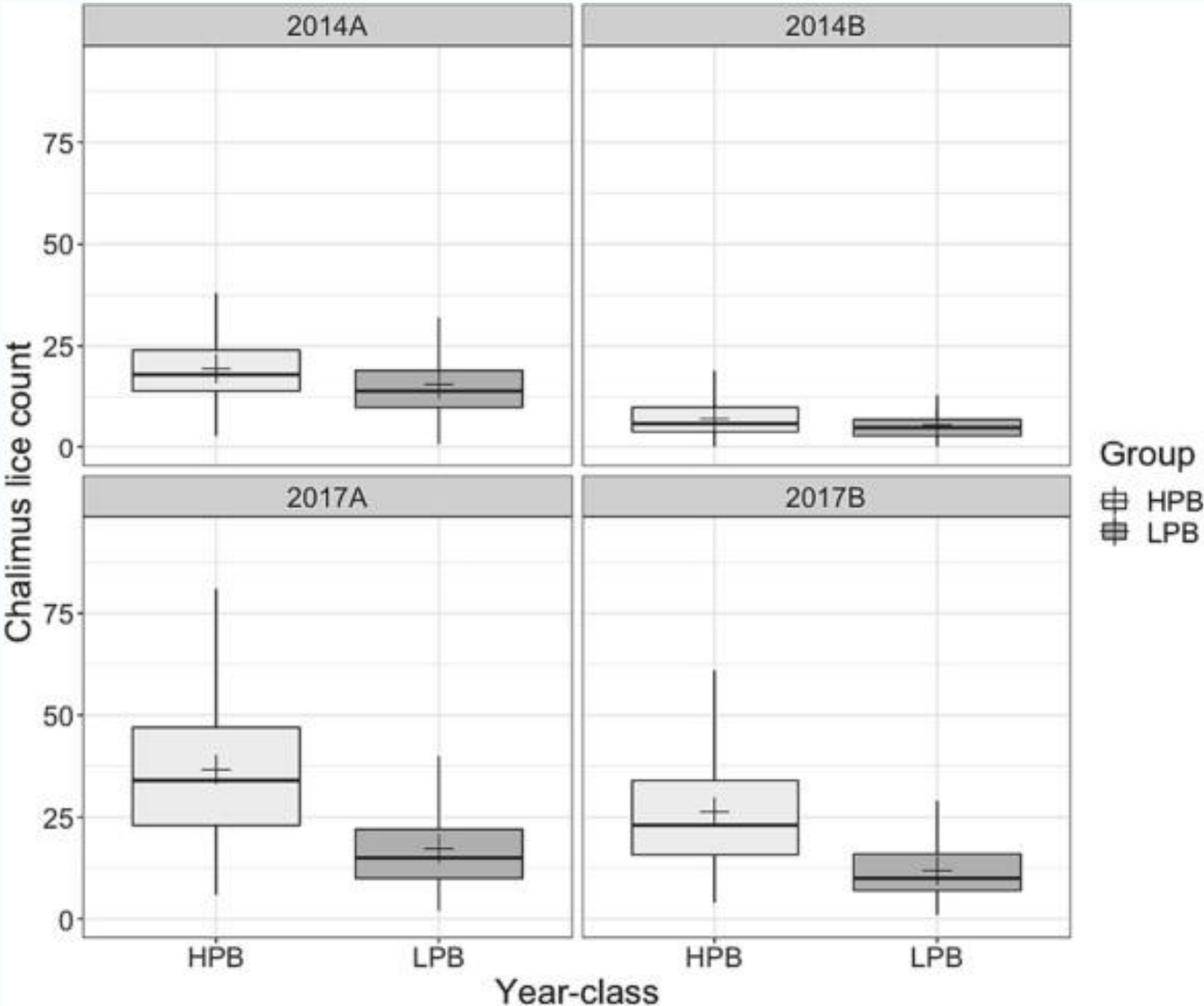


Genomics vs pedigree

Up to 30% increase in survival (Gen 5)

~ 60% more ROI per generation

GENOMIC SELECTION FOR SEA LICE BURDEN

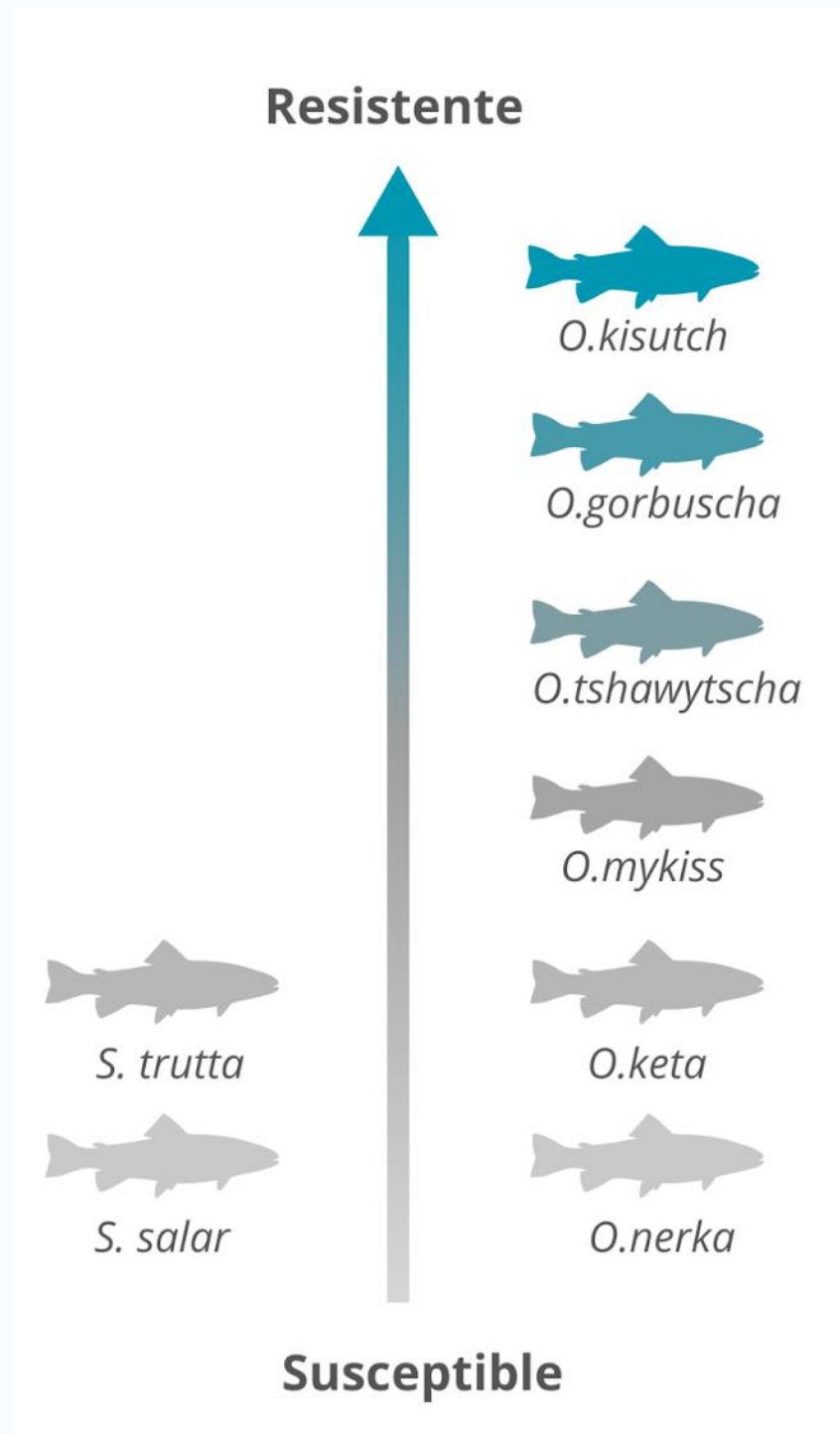


Genetic selection for reduced parasite load in Atlantic salmon: Zero-sum game or a tool for group-level protection against sea lice?

Jørgen Ødegård ^a ✉, Matias Medina ^a, Jacob S. Torgersen ^a, Sven A. Korsvoll ^a, Robert Deerenberg ^b, José M. Yáñez ^{c d}, Daniela Cichero ^b, Paulina Lopez ^b, Thomas Moen ^a, Sissel Kjølglum ^a

Slight or null differences between HPB and LPB when challenged in separate tanks

SEA LICE RESISTANCE: COHO VS SALAR



Salisbury et al. *BMC Biology* (2024) 22:160
<https://doi.org/10.1186/s12915-024-01952-8>

BMC Biology

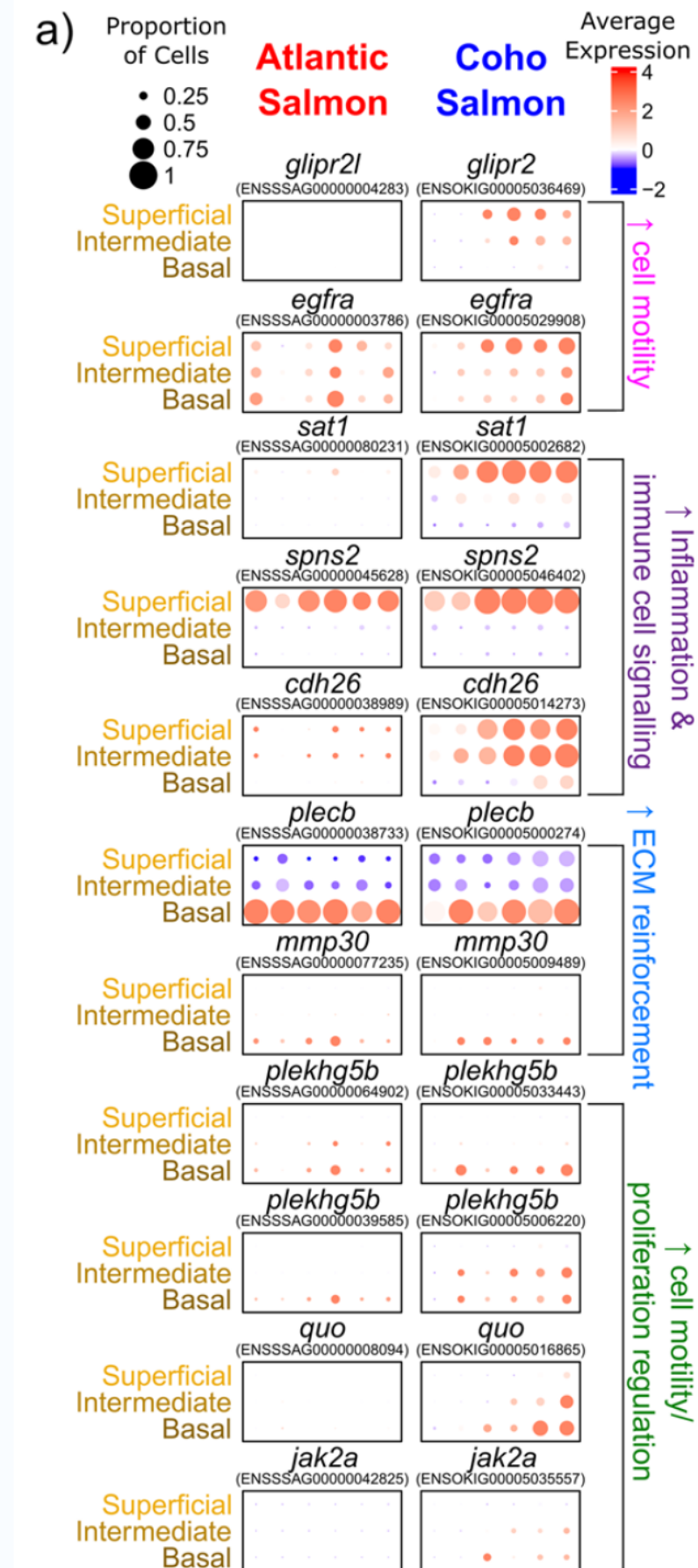
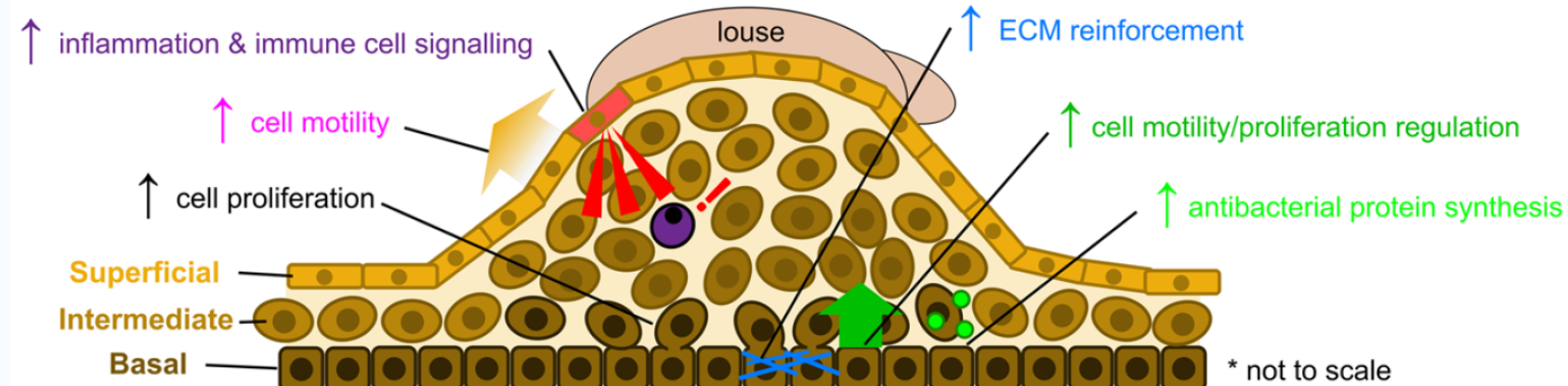
RESEARCH ARTICLE

Open Access



Keratinocytes drive the epithelial hyperplasia key to sea lice resistance in coho salmon

S. J. Salisbury^{1*}, R. Ruiz Daniels^{1†}, S. J. Monaghan², J. E. Bron², P. R. Villamayor^{1,3}, O. Gervais¹, M. D. Fast⁴, L. Sveen⁵, R. D. Houston⁶, N. Robinson^{5,7*} and D. Robledo^{1,3*}

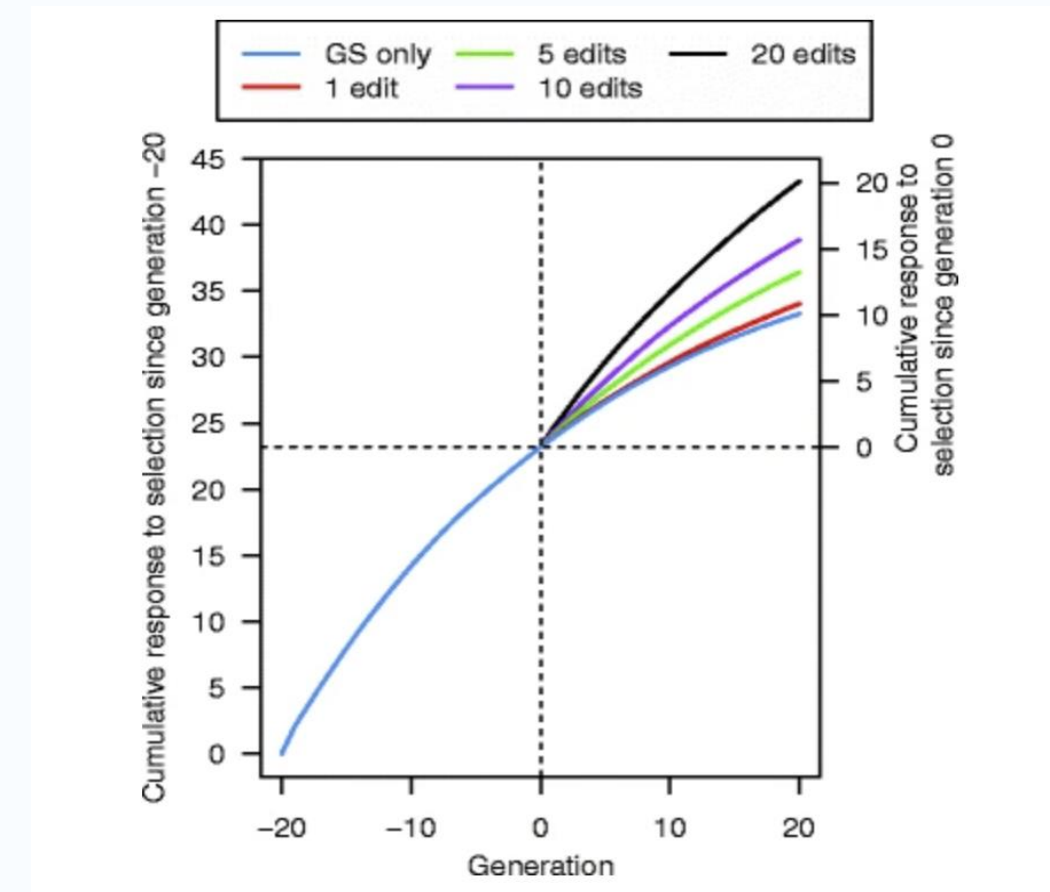




GENE EDITING (CRISPR/CAS9)

EDIGEN

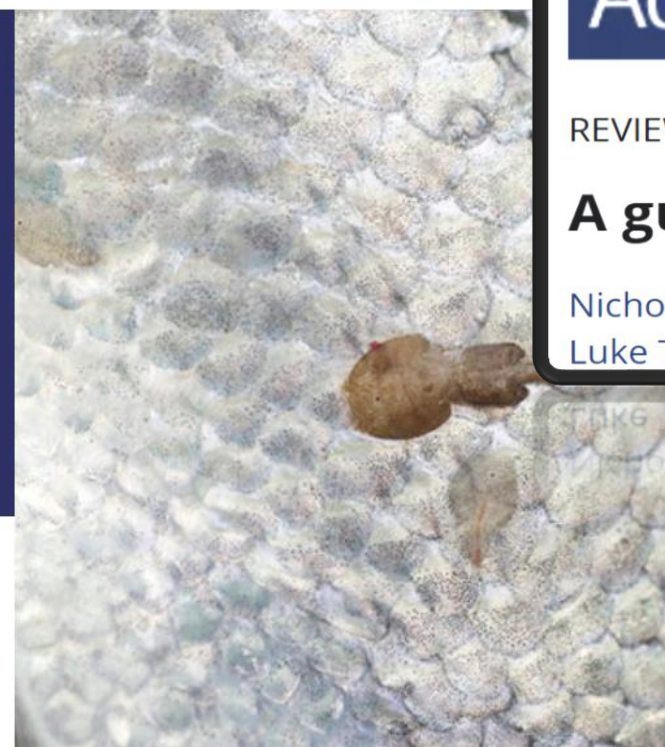
- Permite modificar el ADN de un organismo
- Más rápido, barato, preciso y eficiente que otros métodos de ingeniería genética
- Puede replicar cambios que podrían ocurrir de forma natural o por selección tradicional
- No necesariamente implica insertar ADN foráneo (**no transgénesis o GMO?**)



CrispResist

Harnessing cross-species variation in sea lice resistance

Do Pacific salmon species carry a secret they could share with Atlantic salmon? What is it in the genes that make these relatives of Atlantic salmon less susceptible to lice? Researchers is looking for the answers to these questions, using the gene editing method CRISPR-Cas9.



Sea lice on salmon. Photo: Helge Skodvin, Nofima.

REVIEWS IN Aquaculture

Reviews in Aquaculture homepage

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A guide to assess the use of gene editing in aquaculture

Nicholas A. Robinson✉, Tone-Kari Knutsdatter Østbye, Anne H. Kettunen, Andrew Coates, Luke T. Barrett, Diego Robledo, Tim Dempster

Date last updated:	12/01/2023	
Updated by:	Nick Robinson	
Project name:	CrispResist	
Aim:	Provide Atlantic salmon with host resistance to sea lice	
Is the ultimate purpose to produce animals for aquaculture?	Yes	Note: If yes, continue with assessment. If the gene editing is for another purpose (e.g. research tool to test the function of genes or for producing a gene drive for pest control) another assessment procedure is needed.
Availability of alternative solutions?	No	Note: If highly effective alternative solutions are available, consider whether gene editing is appropriate
Strategy for rolling out gene editing	Introduce at multiplier level by injection of fish	Note: A detailed strategy for implementation is essential

b)

TOPIC			BENEFIT ASSESSMENT			
ID	Area of concern	Weight (1-10)	Notes on potential benefits	Probability of positive impact	Magnitude of benefit	Benefit rating
3	Effect on disease risk for cultured and wild pops		Resistance in farmed salmon could act like a vaccine affecting the prevalence and transmission of infection to wild populations of salmon (herd effect).	High	High	Highly beneficial

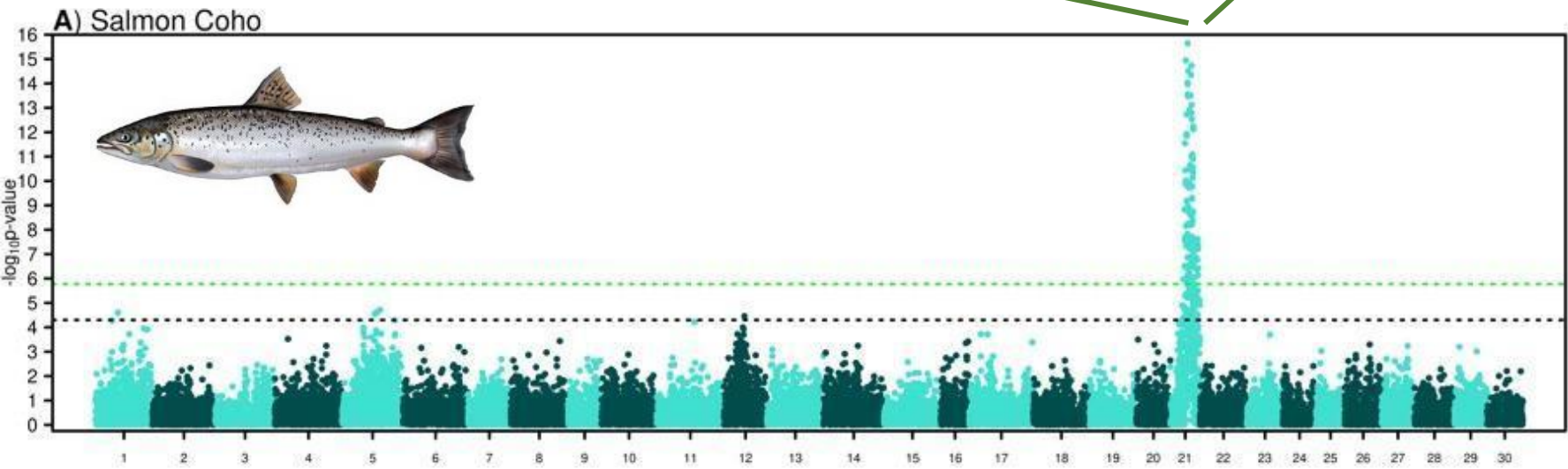
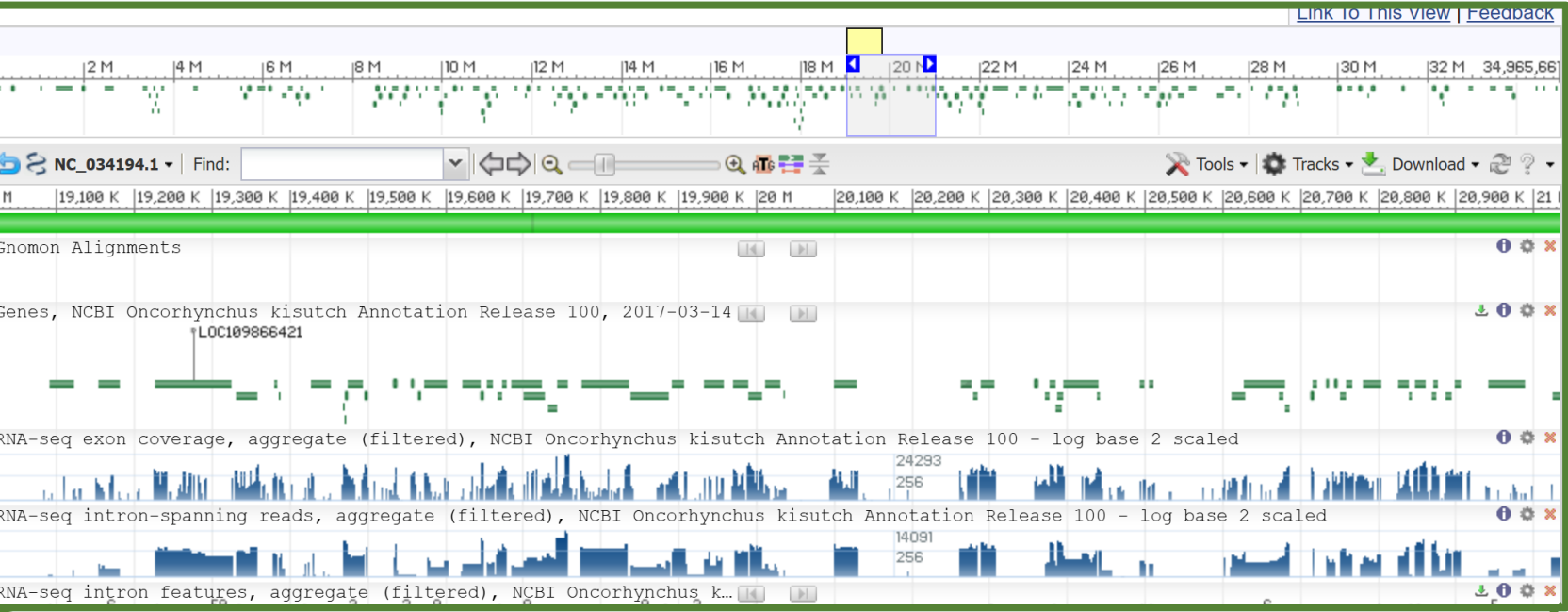
c)

RISK ASSESSMENT									OUTCOME
Notes on potential harms	Probability of negative impact	Magnitude of harm	Initial risk rating	Risk mitigation or control (s)	Control owner (s) name and role	Residual probability of negative impact	Residual magnitude of harm	Residual risk rating	Net benefit score
Parasite or pathogen evolves to overcome resistance, possibly becoming more virulent.	Medium	High	Severe	Design edit in such a way as it is difficult to counter, introduce multiple edits affecting the trait and deploy in time and space, and alongside other preventative methods, in ways that slow evolution of the parasite.		Very low	High	Moderate	2

Risks versus Benefits



CHILE: SRS RESISTANCE



Functional validation of quantitative trait loci
controlling resistance to Salmon Rickettsial
Syndrome in coho salmon (*Oncorhynchus kisutch*)



In vitro Gene Editing for SRS resistance
genes in cell cultures

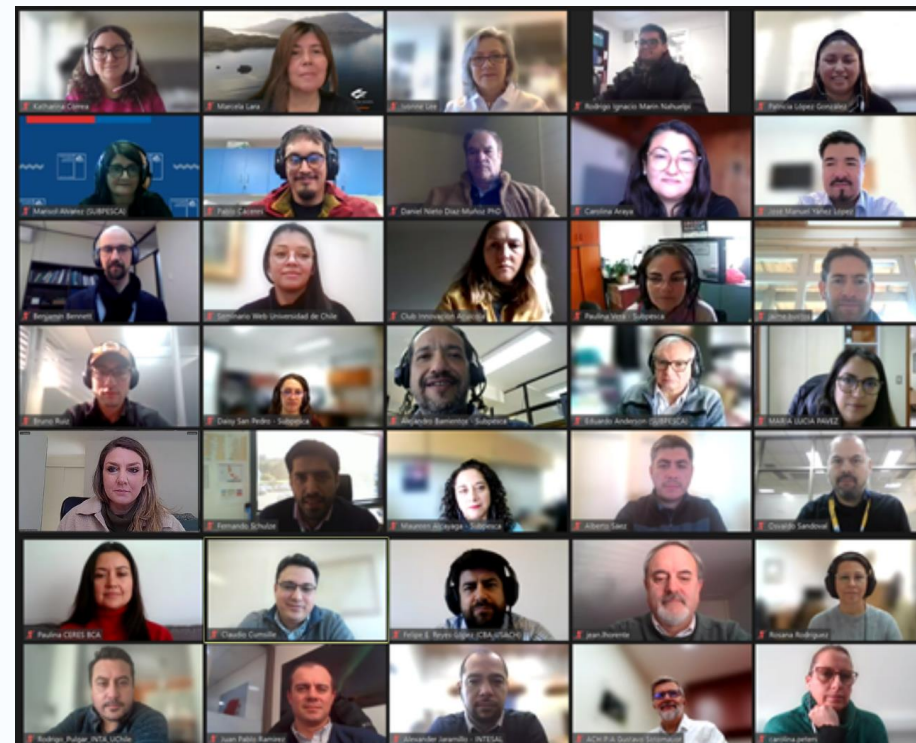


EDIGEN

Iniciativa público-privada que abre un debate amplio e informado sobre la viabilidad de aplicar **edición genética** en animales acuáticos desde una perspectiva científica, normativa y comercial

Objetivo

Generar estrategias de apoyo a la interpretación de la normativa vigente para impulsar la investigación, competitividad y sostenibilidad de la industria, manteniéndola alineada con las tendencias globales



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SAVE THE

19th - 20th Nov, 2025 DATE

**First International Meeting on
Genome Editing in Aquaculture**

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Primer Seminario Internacional de Edición Genética en Acuicultura



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Nofima, Noruega.
Universidad Deakin, Australia.



Derek Bickhart, Ph.D.
Hendrix Genetics, Países Bajos.



Ross Houston, Ph.D.
Benchmark Genetics, Reino Unido.



Miguel Allende-Conelly, Ph.D.
Instituto Milenio Centro de Regulación del Genoma,
Universidad de Chile.



November 19th – 20th, 2025



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Attending and online formats



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El desempeño de una ova no es universal:
depende del ambiente y de las condiciones productivas

Diversificar el origen genético fortalece la resiliencia, asegura la continuidad operativa y mantiene la capacidad de innovación de la industria